DEPARTMENT OF MICROBIOLOGY, OSMANIA UNIVERSITY M.Sc. (Previous) MICROBIOLOGY (Semester System) Syllabus, Scheme of Instruction and Examination

Semester I (15 Weeks)

Paper No.	Sub.Code	Subject/Paper Title		Instruction Hrs/Wk	Exam Time Hrs	Max Marks
		THEORY				
I	MB 101	General Microbiology		4	3	100
II	MB 102	Virology		4	3	100
III	MB 103	Biometry and Computers		4	3	100
IV	MB 104	Biochemistry		4	3	100
		PRACTICALS				
I	MB 151	General Microbiology		4	a	-
II	MB 152	Virology		4	b	-
III	MB 153	Biometry and Computers		4	c	-
IV	MB 154	Biochemistry		4	d	-
	MB 199	Seminar		4	-	-
			Total	32+4		400

Semester II (15 Weeks)

Paper No.	Sub.Code	Subject/Paper Title	Instruction Hrs/Wk	Exam Time Hrs	Max Marks
		THEORY			
I	MB 201	Microbial Physiology	4	3	100
II	MB 202	Immunology	4	3	100
III	MB 203	Antimicrobial Agents and Chemotherap	y 4	3	100
IV	MB 204	Enzymology and Biochemical Technique	ies 4	3	100
ī	MB 251	PRACTICALS Microbial Physiology	4	6(a)	100
II	MB 252	Immunology	4	6(b)	100
III	MB 253	Antimicrobial Agents and Chemotherap	=	6(c)	100
IV	MB 254	Enzymology and Biochemical Technique	•	6(d)	100
	MB 299	Seminar	4	-	-
		Total	32+4		400

Note: (a) The examination includes both MB 151 and MB 251 syllabi

- (b) The examination includes both MB 152 and MB 252 syllabi
- (c) The examination includes both MB 153 and MB 253 syllabi
- (d) The examination includes both MB 154 and MB 254 syllabi

M.Sc. (Previous) I Semester Paper I MB 101 General Microbiology (4 Units x 15 Hrs = 60 Hrs teaching)

Unit I

History of Microbiology. Microscopy. Structure of microbial cells: Spontaneous generation and germ theory of disease, Contributions of Antony van Leeuwenhock, Louis Pasteur, Robert Koch, Edward Jenner, Winogradsky, Beijerink, Alexander Flemming, Waksman. Developments in modern biology.

Principles and working of bright field microscope, fluorescent microscope, phase contrast microscope, electron microscope. Application and importance of above microscopes. Measurement of microscopic objects. Prokaryotic cell, Eukaryotic cell, Organization and function of cellular organells

Cell wall of bacteria, cell membranes, flagella, pili, capsule structure, chemical structure of peptidoglycan, protoplasts, spheroplasts, microsomes and ribosomal RNAs, Nuclear material/nucleus.

Unit II

Methods of sterilization: Physical methods – Dry heat, moist heat, radiation methods, filtration methods, chemical methods and their application. Concept of containment facility, sterilization at industrial level

Microbial cultures: Concept of pure culture, Methods of pure culture isolation, Enrichment culturing techniques, single cell isolation, and pure culture development.

Preservation and Maintenance of Microbial cultures: Repeated subculturing, preservation at low temperature, sterile soil preservation, mineral oil preservation, deep freezing and liquid nitrogen preservation, freze-drying (lyophilization). Advantages and disadvantages of each method.

Unit III

Identification methods and classification of bacteria: -

Microscopic identification characteristics, staining methods – simple staining, differential staining, structural staining and special staining methods.

Ecological identification methods, Nutritional (cultural) identification characters, chemical identification characters, biochemical identification methods, immunological characteristics, pathogenic properties identification, genetic characteristics identification.

Principles of bacterial taxonomy and classification: - Numerical taxonomy, Bergy's manual and its importance, general properties of bacterial groups.

Rickettsiae-General characters and their importance; Mycoplasma – general characters; Chlamydiae – TRIC agents and LGV

Unit IV

Algae, Fungi and their characters:

Distribution of algae, thallus organization, products of algae and their importance.

Reproduction, Biochemical classification of algae.

Vegetative body of fungi, Reproduction, fruiting bodies and dispersal of fungal propagules. Nutritional groups and habitat relationships of fungi. Economic importance of fungi. Classification of fungi.

I Semester MB 151 General Microbiology (Practicals)

General instructions, Microbiology laboratory and its discipline
Handling of microscopes, Calibration and measurement of microscopic objects
Staining techniques for bacteria – simple, differential and special stainings
Preparation of media and reagents/stains
Sterilization procedures/methods
Isolation and cultivation of pure cultures
Identification methods of bacteria
Isolation and culturing of fungi (yeasts and molds) and algae
Observation of specimen and permanent slides

Recommended books

Text book of Microbiology by M. Burrows

General Microbiology by Stainier, Deudroff and Adelberg

Review of medical microbiology by Jawitz, melnick and Adelberg

Bacterial and Mycotic infections of man. Ed. Dubos and Hirst Lipincott

Principles of Microbiology and Immunology by Davis, Dulbecco, Eison, Ginsberg and Wood.

Microbiology by Pelczar M.J., Ried, RD and Chan, ECS.

Structure and Reproduction of Algae, Vol. I & II by Fritsch, F.E.

Introduction to Algae by Morris, I.

Products and Properties of Algae by Zizac.

Fresh water algae of the United States by Smith, GM.

Introductory Mycology, by Alexopolus, C.J.

Dispersal in Fungi by Ingold, CT

Microbial Physiology by Moat,

Microbes in Action by Seoley HW and Van-Demark, PJ

Laboratory Experiments in Microbiology by Gopal Reddy et al

Brock's Biology of microorganisms by Madigan, MT et al

M.Sc. (Previous) I Semester Paper II MB 102 Virology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Classification of viruses, Methods of cultivation, detection, quantitation. Propagation and maintenance of viruses (bacterial, plant and animal viruses). Structure and replication of plant viruses (a) TMV (b) Cauliflower mosaic virus

Unit II

Structure and replication of bacteriophages: Lytic ds linear DNA viruses (T2, T7); Lysogenic ds linear DNA virus (Lambda); ss Circular DNA virus (¢ X 174); Male specific filamentous ss RNA virus (F17 and M13)

Unit III

Recombination in phage, multiplicity reactivation and phenotypic mixing. Structure and replication of animal viruses (Adenovirus (eg. Type 2)

Unit IV

Structure and r eplication of Myxoviruses (eg. Influenza); Pox virus (eg. Vaccinia); Hepatitis virus.

Effect of animal virus infection on host cell; Viral interference and interferon; Tumor viruses (DNA and RNA)

I Semester Paper II MB 152 Virology (Practicals)

Isolation of phage from soil.

Cultivation and preservation of phages

Ouantitation of phages

Growth phages of phage and burst size

Isolation of plaque type and host range mutants

Phage induction

Lysogeny-Transduction

Propagation of animal viruses in egg allantoic, amniotic and CAM

Propagation of viruses in tissue culture

Plaque titration and neutralization

Quantitation of virus by HA, Pock titration, plaque titration

Identification of virus – HAI

Demonstration of cytopathological changes

Symptomatic observations of plant viral infections

Chloroplast agglutination of plant viruses

Recommended Books

General Virology by Luria and Darnel
Virology and Immunology by Jokli
Text book of Virology by Rhodes and Van Royen
Plant Virology by Smith
Genetics of bacteria and their viruses by W. Hayes
Molecular Biology of the gene by Watson, Roberts, Staitz and Weiner
A laboratgory guide in virology by Chjarles H. Lunningham
Basic lab procedures in diagnostic virology by Marty Cristensen

M.Sc. (Previous) I Semester Paper III MB 103 Biometry and Computers (4 units x 15 hrs = 60 hrs teaching)

Unit I

Introduction to statistics and biometry

Population, samples and sampling procedures, variables, variations and frequency distributions, measures of averages and dispersion, element of probability, gausian or normal distribution, binomial distribution, poisson distribution, 't' distribution, 'F' distribution and Chi-square distribution, correlation and linear regression with a special reference to applications in biology

Unit II

Tests of inference and concepts

Small sample tests and large sample tests: Normal curve test, 't' test, 'F' test, ANOVA, analysis of covariance, Chi-square test, and confidence intervals

Methods of collection of data, experimental designs and sampling procedures

Growth curves – exponential and linear growth curves

Example oriented syllabus with applications to biology and microbiology

Unit III

Introduction to Computers

Basic concepts (input, output, CPU, ALU etc.), what is Hardware and software, brief introduction on input output devices (Disks, Printers, CD-ROMS and other storage media etc.)

Introduction to disk operating systems (DOS): Sample commands, DIR-CD-RD-DEL-COPY-MOVE-REN-TYPE-EDIT (Editor) CE-DATE and TIME.

Introduction to Windows 95: Icons, Desktop, Files and Folders.

Simple operations like creation, deletion, moving, copying of Files or folders using windows explorer. Searching files and folders. Creating shortcuts and other simple operations.

Unit IV

Word Processing: Opening, creating and saving documents – Typing, navigating and selecting – editing and sorting-checking spelling and grammar – formatting – changing appearance of your page – importing graphics – working with tables – working with land documents – Printing

Electronic Spread Sheet: Creating, opening and saving files – working with worksheets and work books – entering data and selecting cells – editing worksheets – printing, creating formulae – working with charts – Summarize data in lists and tables – Analyzing data with Pivot tables – Analyzing sample statistical data – Validating cells.

I Semester Paper III MB 153 Biometry and Computers (Practicals)

Formation of frequency distribution, graphical presentation and calculation of descriptive measures

Fitting of distribution – Binomial, Poisson and Normal distribution Chi-square test, goodness of fit, contingency and heterogeneity Small and large sample tests, sample means and properties Problems on correlation and regression ANOVA – One way and two way classifications Creating documents using word processor Exercising the features of word processor Creating spread sheets

Usage of spread sheet to biological applications

Recommended Books

Introduction to the theory of statistics by Alexander, M Mood and Franklin Fundamentsls of Biometry by L.N.Balam Statistical methods by Snedecor and Cochran Introduction to computer and its application by Chae C.Chien Basic Programming language by Bajaraman Biostatistics – A manual of statistical methods for use in Health, Nutrition and Anthropology by K. Vishveshwar Rao

M.Sc. (Previous) I Semester Paper IV MB 104 Biochemistry (4 units x 15 hrs = 60 hrs teaching)

Unit I

pH and its biological relevance Determination of pH, preparation of buffers Concept of entropy, free-energy, free energy changes, high energy compounds. Equilibrium constants, Redox potentials, Biological redox systems

Unit II

Biological oxidation, Biological redox carriers, biological membranes, electron transport, oxidative phosphorylation and mechanism. Bacterial photosynthesis, photosynthetic electron transport

Unit III

Lipids claffication: Bacterial lipids, prostaglandins, structure, function, Major steroids of biological importance.

Nucleic acids: Structure and properties of purines, pyrimidines, nucleosides and nucleotides

Metabolism of purines and pyrimidines - Biosynthesis and degradation

Unit IV

Proteins and amino acids: Properties of amino acids, structure, confirmation and properties of proteins, metabolism of amino acids, biosynthesis and degradation – an overvies; Urea cycle

I Semester Paper IV MB 154 Biochemistry (Practicals)

Preparation of buffers and adjustment of pH Qualitative tests for carbohydrates and analysis of unknowns Qualitative tests for amino acids and analysis of unknowns Tests for lipids (qualitative) Quantitative estimation of glucose and fructose

Recommended Books

Biochemistry by Lehninger
Outlines of Biochemistry by Cohn and Stumph
Biochemistry of Nucleic acids by Davidson
Biological Chemistry by Mullar and Cards
Biochemistry by White, Handler and Smith
Methods in Enzymology series
The Cell – Bratch amd Mirsky series
Laboratory experiments in Microbiology by Gopal Reddy et al
Biochemistry lab manual by Jayaraman

M.Sc. (Previous) II Semester Paper I MB 201 Microbial Physiology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Microbial nutrition – Elemental nutrient requirements of microbes, nutritional groups of bacteria. The autotrophy – Photoautotrophy and bacterial photosynthesis Chemoautotrophy and autotrophic metabolism.

Concept of heterotrophy – Photoheterotrophy and chemoheterotrophy. Heterotrophic metabolism in bacteria. Respiration (Aerobic and anaerobic) and fermentation

Unit II

Microbiological media and culturing and cultivation of microorganisms - Autotrophic media, defined synthetic mineral media, heterotrophic media. The concept of prototrophs and auxotrophs, prototrophic (minimal) media (defined media), complex media (undefined media), Basal medium, enriched media, enrichment media, selective media, biochemical media, differential media, maintenance media, transport media. Media for cultivation of fungi, and algae

Cultivation methods of bacteria, slant culturing, stab culturing, agar plate culturing, rolled tube/bottle culturing, tube cultures, flask culturing.

Aerobic culturing methods, anaerobic culturing methods Environmental requirements of growth

Unit III

Microbial growth: The concept of growth and definition, formation of protoplasm, building of macromolecules from elemental nutrients, supramolecules, orgnelles of cell and cellular components. Cell cycle in microbes and generation time Growth phases of bacteria – Lag phase, exponential (logarithmic) phase, stationary (ideo) phase, decline and survival of microbial cells. Importance of each growth phase. Synchronous cultures – methods of synchronous culturing Continuous culturing methods, factors effecting growth. Methods of growth measurement.

Unit IV

Nature and properties of spores: Bacterial endospore structure, phenomenon of sporulation, biochemistry and genetics of sporulation. Induction of sporulation phenomenon. Germination of spores

The concept of disease: Infectious disease, pathogenic microbes, properties of pathogenic microbes, Infection, pathogenesis and virulence. Virulence factore – Capsular materials, bacterial toxins – exotoxins, toxoids, endotoxins, enterotoxins. Physiology of toxin production. Extracellular enzymes of pathogenic bacteria. Application of toxins and toxic enzymes, laboratory testing methods for bacterial virulence properties

II Semester Paper I MB 251 Microbial Physiology (Practicals)

Preparation of microbiological media. Autotrophic media, minimal media, basic media, enriched media, enrichment media, differential media.

Isolation and cultivation of autotrophic microbes

Culturing methods of microbes – slant and stab cultures, tube culture, flask cultures, shake flask cultures

Anaerobic culturing methods – anaerobic jar and its use, pyrogallol method, thioglycollate media culturing, anaerobic glove box and its application

Microbial growth experiments – Viable count of growing cultures and generation time determination

Determination of microbial growth by turbidometric methods

Study of bacterial growth curve

Factures effecting the microbial growth

Tests for microbial toxins – determination of LD50

Testing for extracellular microbial enzymes

Recommended Books

Text book of Microbiology by M. Burrows

General Microbiology by Stainier, Deudroff and Adelberg

Review of medical microbiology by Jawitz, melnick and Adelberg

Bacterial and Mycotic infections of man. Ed. Dubos and Hirst Lipincott

Principles of Microbiology and Immunology by Davis, Dulbecco, Eison, Ginsberg and Wood.

Text book of Microbiology by Ananthanarayanan

Microbiology by Pelczar M.J., Ried, RD and Chan, ECS.

Microbial Physiology by Moat,

Brock's Biology of microorganisms by Madigan, MT et al

Biochemistry of bacterial growth by Mandelstum, Mc Quillon and Dawes

Bacterial Metabolism by Dwellely

Photosynthesis by Dewlin and Barker

Laboratory Experiments in Microbiology by Gopal Reddy et al

Microbes in Action by Seoley HW and Van-Demark, PJ

M.Sc .(Previous) II Semester Paper II MB 202 Immunology (4 units X 15 hrs=60 hrs teaching)

Unit I

Brief history of immunology; Antigens – isoantigens, alloantigens, haptens; Antibodies – structures, genes; Structural variants, classes, and subclasses of antibodies; Immune system – cells, vessels, organs; Immune response – antigen recognition; T-cell interactions; B-cell interactions.

Unit II

Immune response; Types of immunity – innate, specific, acquired, prophylaxis; Vaccines – whole cell, subunit, recombinant protein-based, DNA-based; Production of vaccines; QA and QC of vaccines; Accepted pharmacopeial techniques; Future of vaccines – bacterial and viral vaccines in market; Advantages and disadvantages of vaccines; Approaches of vaccine design; Acquired and natural tolerance; Mechanism of tolerance; Properties of interferon; Production and assay of interferons.

Unit III

MHC Complex; Immunoglobulin superfamily; Self and non-self recognition; Genetics of tissue transplantation; Genetic control of immune response; Macrophage restriction; Structure of MHC; MHC products and cell surface; MHC and tissue transplantation; Human MHC and disease association; Autoimmunity and HLA; Examples of autoimmunity and treatment; Inflammation and anaphylactic reactions; Immune reaction of infection, reactions to infectious agents.

Unit IV

Antigen-antibody reactions; Polyfunctional antibodies; Cross reactivity; Primary antigenantibody reactions *in vivo*; Secondary antigen-antibody reactions *in vivo*; Sensitivity of antigen-antibody reactions; Immunological methods – agglutination, precipitation, complement fixation, radioactive and enzyme-mediated amplification for detection of antigen-antibody reactions; Immunization – adjuvants, lymphokines; Alternate pathway for complement fixation; Protective immunity to infection; Role of immune mechanism in protection against infection; Immune effector mechanisms in specific infections; Evasion of immune defenses; Tumor immunity – host-tumor relationship, tumor immunity in humans; Tumor antigens; Mechanism of tumor destruction by the immune system; Tumor surveillance; Immuno-diagnosis or cancers; Immuno-prophylaxis of cancer; Immuno-therapy of cancer.

II Semester Paper II MB 252 Immunology (Practicals)

Agglutination reactions – Widal, VDRL, HA, Blood group/typing Precipitation test: Ring interphase, single radial diffusion, Ouechterlony, immunoelectrophoresis

Neutralization test – Plaque neutralization, Haeme absorption test Raising of antisera

Seperation of serum, WBC, RBC, Plasma, CBP and differential blood picture Lymphoblast transformation, Jerme plaque test, T-cell migration.

Seperation of serum proteins

Blot transfer and detection of protein on blot by staining

Lymphocyte viability test

ELIS, Indirect agglutination (a) Hepatitis (b) Pregnancy hCG Ag Raising of polyclonal Ab and Monoclonal Ab

Recommended Books

Immunology and immunopathology by Stewart Sell
Cellular and molecular immunology by Abul K. Abbas et al
Immunology by Herman N. Eosen
Test book of Immunology by Barret
Molecular basis of immunology by Constantin Bena
Immunology – The science of self-non self discrimination by Jan Klein
Essential Immunology by Roitt, IM
Immunology by Kuby, J.

M.Sc. (Previous) II Semester Paper III MB 203 Antimicrobial Agents and Chemotherapy (4 units x 15 hrs = 60 hrs teaching)

Unit I

History of chemotherapy. Types of antimicrobial agents, Plants and arsenicals as therapeutic agents, Paul Ehrlich and his contributions to chemotherapy. Development of synthetic drugs, development of antibiotics,

Chemical non-medicinal antimicrobials- sanitizers, disinfectants, aantiseptics.

Selective toxicity and target sites of drug action.

Synthetic medicinal agents (drugs)- sulphonamides, antitubercular compounds, nitrofurans, nalidixic acid, metronidazoles,

Antibiotics – Definition of antibiotics, types (chemical) of antibiotics, cell wall inhibitors, membrane inhibitors, inhibitors of macromolelcular synthesis, antimetabolites.

Unit II

Bactericidal and bactedriostatic agents, Factors affecting static and cidal activity, phenols and phenolic compounds, alcohols, halogens, heavy metals, dyes, detergents, aldehydes

Non-medical uses of antibiotics.

Assay methods of antimicrobial agents – Phenol coefficient, qualitative assay of drugs (drug sensitivity testing), quantitative assays – liquid tube assay (MIC), agar tube assay. Agar plate assay.

Unit III

Principles of chemotherapy and Drug Resistance

The physical and lab diagnosis, tests for sensitivity, choice drug determination, dosage, route of administration, combined drug therapy. Policies of antibiotic usage. Antiviral agents – Biological antiviral agents- interferon and its action, chemical antiviral agents. Phenomenon of drug resistance, basis of drug resistance, biochemistry of drug resistance, genetics of drug resistance. Control of drug resistant bacteria

Unit IV

Mode of action of important drugs – Cell wall inhibitors (betalactam drugs), membrane inhibitors (polymyxin), Ribosomal inhibitors (aminoglycosides – streptomycin), folic acid inhibitors (sulfa drugs), antifungal drugs (nystatin).

II Semester Paper III MB 253 Antimicrobial agents and Chemotherapy (Practicals)

Testing for drug sensitivity
Determination of MIC values (tube dilution method)
Determination of phenol coefficient (Reidel Walker coefficient, RWC)
Study for antimicrobial spectrum of antimicrobials
Determination of cidal and static activity
Screening for antibiotic producing microbes
Chemical assay methods for antimicrobial drugs
Microbiological assays of antimicrobial drugs

Recommended books

Disinfection, sterilization and preservation Ed. Block, SS.
Microbial contamaination control facilities Ed. Runkle, RS and Philips, GB
Principles and methods of sterilization in health science by Perkins John, J.
Inhibition and destruction of the microbial cell Ed. Hugo
Biochemistry of antimicrobial action by Franklin, TJ and Snow, L.
The Molecular basis of antibiotic action by Gale et al
Antibiotics and chemotherapy by Gerrod et al
Pharmaceutical microbiologoy by Hugo Russell
Microbiological assays by Hewitt
Antiviral drugs by S.Karger
Burgers' medicinal chemistry (all volumes) Ed. Manfield E. World.
Antibiotic interactions by Williams

The control of antibiotic resistant bacteria by Harris and Harris

M.Sc. (Previous) II Semester Paper IV MB 204 Enzymology and Biochemical Techniques (4 units x 15 hrs = 60 hrs teaching)

Unit I

Enzymes nomenclature, classification methods for determination of enzyme activity. Isolation and purification of enzymes. Enzyme kinetics: Effect of pH, substrate concentration, temperature and inhibitors. Isoenzymes. Competitive and non-competitive inhibition

Unit II

Mechanism of enzyme action – Action of Ribonuclease, chymotrypsin, and trypsin. Coenzyme catalysis. Mechanism of PAXPO and thiamine pyrophosphate enzymes Control of enzymes. Regulation of enzyme activity: allosteric enzymes and feed back mechanisms. Metabolic compartamentalization in relation to enzyme, enzymes and secondary metabolites

Unit III

Optical methods: Elementary treatment of biochemical methods, colourimetry and spectrophotometry, flurometry, optical rotation, fluorescence
Seperation methods: Chromatographic techniques – paper, thin layer, ion exchange, gel filtration and affinity chromatography. Diffusion, dialysis, cell disruption methods, centrifugation techniques, cell free extracts and their use in metabolic studies.

Unit IV

General method of study of intermediary metabolism

Electrophoretic techniques and application, counter current distribution

Radio isotopes – detection and measurement of radioactivity – scientilation counters, autoradiography, stable isotopes and their use. Uses of mutants in study of metabolism

II Semester Paper IV MB 254 Enzymology and Biochemical Techniques (Practicals)

Determination of saponification value and iodine number of fats
Quantitative estimation of inorganic and organic phosphate
Partial purification of enzymes (B-amylase, urease, pyrophosphotase and catalase)
Effect of substrate concentration, pH, time and temperature on enzyme activity
Calculation of Km for partially purified enzyme
Inhibition of enzyme activity

Recommended books

Biochemistry by Lehninger
Outlines of Biochemistry by Cohn and Stumph
Biochemistry of Nucleic acids by Davidson
Biological Chemistry by Mullar and Cards
Biochemistry by White, Handler and Smith
Methods in Enzymology series
The Cell – Bratch amd Mirsky series
Laboratory experiments in Microbiology by Gopal Reddy et al
Biochemistry lab manual by Jayaraman

DEPARTMENT OF MICROBIOLOGY, OSMANIA UNIVERSITY M.Sc. (Final) MICROBIOLOGY (Semester System) Syllabus, Scheme of Instruction and Examination

Semester III (15 Weeks)

Paper No.	Sub.Code	Subject/Paper Title	Instruction Hrs/Wk	Exam Time Hrs	Max Marks
		THEORY			
I	MB 301	Molecular Biology and Microbial Genetic	es 4	3	100
II	MB 302	Industrial Microbiology	4	3	100
III	MB 303	Soil Microbiology	4	3	100
IV	MB 304	Medical Bacteriology	4	3	100
		PRACTICALS			
I	MB 351	Molecular Biology and Microbial Genetic	es 4	a	-
II	MB 352	Industrial Microbiology	4	b	-
III	MB 353	Soil Microbiology	4	c	-
IV	MB 354	Medical Bacteriology	4	d	-
	MB 399	Seminar	4	-	-
		Total	32+4		400

Semester IV (15 Weeks)

Paper No.	Sub.Code	Subject/Paper Title	Instruction Hrs/Wk	Exam Time Hrs	Max Marks
		THEORY			
I	MB 401	Molecular Biotechnology	4	3	100
II	MB 402	Microbial Biotechnology	4	3	100
III	MB 403	Microbiology of Food and Environmen	t 4	3	100
IV	MB 404	Medical Virology and Parasitology	4	3	100
		PRACTICALS			
I	MB 451	Molecular Biotechnology	4	6(a)	100
II	MB 452	Microbial Biotechnology	4	6(b)	100
III	MB 453	Microbiology of Food and Environment	4	6(c)	100
IV	MB 454	Medical Virology and Parasitology	4	6(d)	100
	MB 499	Seminar	4	-	-
		 Total	32+4		400

Note The students of M.Sc Final Microbiology have to undergo an educational tour/training/visit to subject based industries/institutions and submit a report as part of their academic curriculum.

- (a) The examination includes both MB 351 and MB 451 syllabi
- (b) The examination includes both MB 352 and MB 452 syllabi
- (c) The examination includes both MB 353 and MB 453 syllabi
- (d) The examination includes both MB 354 and MB 454 syllabi

M.Sc. (Final) III Semester Microbiology Paper I MB 301 Molecular Biology and Microbial Genetics (4 units x 15 hrs = 60 hrs teaching)

Unit I

Structure of DNA and its biosynthesis – Detailed structure of DNA, variation from Watson and Crick model, Z-DNA, A & B DNA, denaturation and melting curves. Genome organization in prokaryotes and eukaryotes

Enzymes involved in replication, step by step process, heteroduplexes,

Modes of DNA replication- Semiconservative and conservative

Mechanisms of DNA replication in E.coli (bi-directional), Mitochondrial (D-loop), Viral DNA (ROLLING CIRCLE), Single stranded DNA phages (¢ X 174),

Eukaryotic telomere and its replication. Selfish DNA, Alu sequences

Unit II

RNA structure and biosynthesis: m-RNA, r-RNA t-RNA structures

Transcription apparatus and proteins involved in transcription

Prokaryotic and eukaryotic transcription. Processing of t-RNA, r-RNA, m-RNA splicing Concept of ribozyme

Genetic code and Wobble hypothesis

Protein synthesis – Translation in Prokaryotes and eukaryotes

Post translational modifications

Unit III

Mutagenesis- Types of mutagens, molecular basis of mutations, analysis of mutations, site directed mutagenesis and reverse genetics

DNA damage and repair mechanisms. Mutagenicity testing using microbial systems Concept of gene – Benzer's fine structure analysis, introns and exons.

Complementation and functional allelism

Unit IV

Bacterial transformation and recombination- Discovery, detection, molecular mechanisms of transformation, transformation methods

Bacterial conjugation – Sex factor in bacteria, F and HFR transfer, mechanism of transfer, linkage mapping, Mechanism of recombination

Bacterial transduction – transduction phenomenon, methods of transduction, cotransduction, generalized, specialized and abortive transduction, sex-ductions, and their applications

Genetics of eukaryotic viruses

Paper I MB 351 Molecular Biology and Microbial Genetics (Practicals)

Extraction/Isolation of DNA (Plasmid DNA AND genomic DNA) (Mini preparation and large preparation

Estimation of DNA, RNA and Protein (colourimetry)

Determination of molecular weight of DNA, resolved on agarose gel electrophoresis

Determination of molecular weight of protein by PAGE

Induction of mutations by physical/chemical mutagens, screening and isolation of mutants, Replica plating technique

Transformation in bacteria

Conjugation in bacteria

Complementation tests in bacteria

Protoplast preparation and regeneration

Recommended books

Molecular biology by David and Freifelder
Microbial genetics by David and Freifelder
Genetics of bacteria and their viruses by William Hayes
Molecular biology of thee gene by Watson et al
The Biochemistry of nucleic acids by Davidson JN
Molecular biotechnology by Primerose
Molecular Biotechnology by Bernard R. Glick and Jack J Pasternak
DNA Microarrays Ed. M. Schena

M.Sc. (Final) III Semester Microbiology Paper II MB 302 Industrial Microbiology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Introduction to industrial microbiology. Definition, scope, history, microorganisms, properties and industrial products

Screening for microbes of industrial importance. Primary screening, screening for amylase, organic acid, antibiotic, amino acid and vitamin producing microorganisms. Secondary screening. Further evaluation of primary isolates

Detection and assay of fermentation products. Physico-chemical methods and biological assays. Fermentation equipment and its use. Design of fermentor, typer of fermentor, agitation, aeration, antifoam, pH and temperature control.

Unit II

Inoculam media, inoculam preparation

Rawmaterials Saccharides, starchy and cellulosic materials

Fermentation media and sterilization.

Types of fermentations processes – Solid state, surface and submerged fermentations

Unit III

Batch, fed batch and continuous fermentations. Direct, dual or multiple fermentations. Scaleup of fermentations. Product recovery methods.

Fermentation type reactions, alcoholic, lactic acid, mixed acid, propionic acid, butandiol and acetone-butanol types

Unit IV

Strain development strategies. Environmental factors and genetic factors for improvement. Immobilization methods – Absorption, covalent linkage, entrapment and cross linkage, types of carriers, advantage and disadvantages.

Paper II MB 352 Industrial Microbiology (Practicals)

Screening for amylase producing organisms
Screening for organic acid producing microorganisms
Isolation of antibiotic producing microorganisms by crowded plate technique
Isolation and culturing of yeasts
Seperation of amino acids by chromatography
Estimation of glucose by DNS method
Estimation of ethanol by dichromarte method
Extimation of maltose
Immobilization of microbial cells by entrapment method

Recommended Books

Industrial Microbiology by Casida, LE
Industrial Microbiology by Patel, AH
Industrial Microbiology by Miller, BM and Litsky
Industrial Microbiology by Prescot and Dunn
Microbial Technology by Peppler, JH and Perlman, D.
Biochemistry of Industrial Microorganisms, by Rainbow and Rose
Economic Microbiology by Rose Vol I – V
Microbial Enzymes and Biotechnology by Fogarty WM and Kelly, CT
Comprehensive Biotechnology, All volumes Ed. Murray Moo-Yong
Biotechnology (A text book of industrial Microbiology) Ed. Cruger & Cruger
Advances in Applied Microbiology Ed. Perlman Series of volumes
Laboratory experiments in microbiology by Gopal Reddy et al

M.Sc. (Final) III Semester Microbiology Paper III MB 303 Soil Microbiology (4 units x 15 hrs = 60 hrs teaching)

Unit I

The soil as habitat for microorganisms: general description of soil, soil structure, differences among soils and factors of ecological significance
Soil Microorganisms: Distribution, abundance, methods of estimation, biomass measurement, environmental factors, activity and functions of soil bacteria, fungi, algae, protozoa, blue green algae and soil fertility
Microbial diversity in soil and its significance

Unit II

Organic matter decomposition both native and added organic matter and factors governing the decomposition

Degradation of carbonaceous materials in soil – cellulose, hemicellulose and lignin decomposition, factors governing the decomposition and biochemistry of decomposition Mineralization of nitrogenous organic matter – microbes involved and factors influencing the processes, Soil humus formation

Unit III

Nitrification – Microbes involved, factors influencing nitrification, nitrifying bacteria and biochemical mechanism. Denitrification – microbes involved, factors influencing and the mechanism of denitrification and nitrate pollution

Nitrogen fixation – Asymbiotic and symbiotic nitrogen fixation, microorganisms involved, biochemistry and genetics of nitrogen fixation, measurement of nitrogen fixation, ecological and economic importance of nitrogen fixation.

Unit IV

Microbial transformation of phosphorus in soil
Microbial transformation sulfur in soil
Microbial transformation iron in soil
Biofertilizedrs – bacterial fertilizers and production of rhizobial inoculants and bluegreen algae, quality control tesrts

Paper III MB 353 Soil Microbiology (Practicals)

Setting of Winogradsky's column

Estimation of soil composition by sedimentation method

Enumeration of soil microorganisms (bacteria), actinomycetes, fungi) by standard plate count

Estimation of soil microbial activity by CO2 evolution

Isolation of cellulose decomposing microbes and estimation of cellulose activity Estimation of ammonifiers, nitrifiers and denitrifiers in soil by MPN METHOD

Isolation and culturing of Rhizobium sp from root nodules and Azospirillum from grasses (Cyanodon)

Biological enrichment isolation of Rhizobium from soil by Leonard Jar experiment

Nodulation testing by tube/jar method

Solubilizatgion of rock phosphate by microorganisms and estimation

Testing for mineral leaching by Thiobacillus sp

Assessment for sulphate reducing bacteria

Observation and assessment of soil algae/algal biofertilizers

Bacterial biofertilizers and their use

Estimation of N2 fixation (Micro Kjedhasl method/GC method)

Recommended Books

Soil Microbiology by Alexander Martin

Microbial ecology, Fundamentals and Applications Ed. Benjamin-Cummings

Soil Biotechnology by JM Iynch

Microbial Ecology: Organisms, Habitats, Activities by Stolp, H.

Soil Microbiology and Biochemistry by Paul E. and PE Clank

Microbial Ecology: Principles, Methods and Applications by Lavin, Seidler, Rogul

Biological Nitrigen Fixation by Quispel

Soil Microorganisms and Plant Growth by N.S,. SubbaRao.

Laboratory experiments in microbiology by Gopal Reddy et al

M.Sc. (Final) III Semester Microbiology Paper IV MB 304 Medical Bacteriology (4 units x 15 hrs = 60 hrs teaching)

Unit 1

Principles of Medical Microbioology:

Classification of medically important microorganisms. Normal flora of human body – Origin of normal flora, role of the resident flora, effect of antimicrobial agents on normal flora, characteristics of normal floraa

Distribution and occurance of normal flora (Skin, conjunctiva, nose, nasopharynx, sinuses, moutn, upper respiratory tract, intestinal tract, urogenital tract)
Bacteria in the blood and tissues, factors sinfluencing normal flora.

Unit II

Properties of pathogenic microorganisms. Factors that influence pathogenicity Type of infections, source of infections, different modes/means of infections

Disgnostic microbiology – Types of specimen, specimen collection, transportation of specimen, processing, laboratory investigations, specific lab. Tests, non-specific lab tests, diagnosis and report

Use of lab animals in diagnostic microbiodlogy

Unit III

Systematic bacteriology – Detailed study of morphology, cultural characteristics, antigenic structure, pathogenesis, diagnostic lab test, epidemiology, prevention and

treatment of the following bacterial pathogens.

Bacterial air borne infections – B-Haemolytic streptococco, pneumococci, Corynebacteium diphtheriae, Mycobacterium tuberculosis, Mycobactterium leprae, Neisseria meningitides, haemophilus influenzae.

Sexually transmitted diseases caused by bacteria, *Treponima pallidum, Neisseria gonorrhoea*

Unit IV

Sysstematic bacteriol9ogy – Detailed study of morphology, cultural characteristics, antigenic structure, pathogenesis, diagnostic lab tests, epidemiology, prevention and treatment of the following pathogenic bacteri

Water borne infections – E.coli, Salmonella typhi, Shigella dysenteriae,s Vibrio choleraae. Wound infections – Staphylococcus aureus, Clostridium tetani, Clostridium welchi, Pseudomonas.

Paper IV MB 354 Medical Bacteriology (Practicals)

Preparation of different types of culture media/observation. Mannitol salt agar, Blair Parker medium, MacConkey agar, Lowensten-Jension medium, Wilson Blair Bismuth sulphite medium, Biochemical media.

Staining techniques – Gram's staining, AFB staining, Albert Staining, Capsular staining Isolation and identification of various pathogenic bacteria by microscopic, biochemical, enzymatic and serological tests

Bacteriological examination of urine, pus, throat swab etc from patients for diagnosis

Recommended Books

Review of Medical Microbiology by Jawitz, Melnick and Adelberg Diagnostic Microbiology by Bailey and Scott Medical Microbiology by Cruckshanak et al Vol I & II Text book of Microbiology by Ananthanarayanan and jayaram Paniker Bacterial diseases by Wilson and Topley Medical Microbiology by Sherries Laboratory experiments in microbiology by Gopal Reddy et al

M.Sc. (Final) IV Semester Microbiology Paper I MB 401 Molecular Biotechnology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Gene regulation and expression – Lac operon, arabinose and tryptophane operons Repressors and activators in lambda, Sigma switch in B.subtilits Gene regulation in eukaryotic systems, repetitive DNA, gene rearrangement, promoters, enhancer elements, gene amplification. Signal transduction – Concept of second messenger, cAMP, cGMP, protein kinases, G-proteins Steroid/peptide hormone regulation, tissue specific regulation.

Unit II

Plasmids – Definition, types of plasmids, identification and classification of plasmids, purification of plasmids, plasmid transfer and its smechanism. Host restriction in transfer. Transposable elements – Definition, detection of transposition in bacteria, types of bacterial transposons, mechanisms of transposition and excision, applications of transposons.

PCR Technology – Principle, designing of primers PCR methodology, RT-PCR, multiplex PCR, identification of PCR products, application of PCR technology.

Unit III

Genetic engineering – Cloning vectors, enzymes involved in genetic engineering, restreiction mapping, cloning strategies, methods of gene transfer.

Detection of clones and their expression – Expression of cloned genes in yeast and E.coli, Blot analysis – Southern, Northern and Westerns blotts, DNA methylation, DNA hybridization.

Genomic and c-DNA library construction and application.

Hybridoma technology – Principle and application

Unit IV

Molecular markers -RFLP, RAPD, AFLP, 16S r-RNA typing.

Gene chip and microarays- Assays, applications in disease profiles, drug target, gene discovery, drug action and toxicity

Molecular biology data bases and bioinformatics – Overview and data mining of sequence data bases. DNA sequencing methodology and soft ware, sequence comparison and alignment. Concept of pharmacogenomics and its application IPR and patenting – Concept and overvies

Paper I MB 451 Molecular Biotechnology (Practicals)

Restriction mapping
PCR technique – demonstration
Plasmid detection and Isolation of Plasmid DNA
Preparation of competent cells and transformation
Gene cloning in bacteria – demonstration

Southern transfer – demonstration
Demonstration of RFLP/AFLP
Induction of enzymes – Lac operon
Transposon mutagenesis
Dialysis and separation of proteins by column chromatography

Recommended Books

Molecular biology by David and Freifelder
Microbial genetics by David and Freifelder
Genetics of bacteria and their viruses by William Hayes
Molecular biology of thee gene by Watson et al
The Biochemistry of nucleic acids by Davidson JN
Molecular biotechnology by Primerose
Molecular Biotechnology by Bernard R. Glick and Jack J Pasternak
DNA Microarrays Ed. M. Schena

M.Sc. (Final) IV Semester Microbiology Paper II MB 402 Microbial Biotechnology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Fermentative production of insustrial alcohol, uses, rawmaterials, microorganisms, inoculum preparation, preparation of wort, fermentation and recovery

Fermentative production of beer – Medium components, malt, malt a djuncts, hops, water. Preparation of wort, mashing, wort boiling, microorganism, inoculum preparation, fermentation, cold storage matduration, carbonation, packing and preservation

Principles of wine making – Fruit selection, picking, crushing, sulphite addition, processing, fermentation, aging and botling

Unit II

Fermentative production of citric acid, uses, microorganism, inoculum preparation, medium preparation, fermentation, recovery and mechanism opf citric acid production Fermentative production of vitamin B12 – Uses, structure of vit.B12, microorganisms, inoculum preparation, medium preparation, fermentation and recovery Fermentative production of glutamic acid – Uses, microorganism, inoculum preparation, production medium, fermentation and down stream processing

Unit III

Antibiotics – Commercial production of benzyl penicillin, uses, microorganism, inoculum preparation, production medium medium, fermentation, recovery and semisynthetic penicillins

Fermentative production of tetracyclines – uses, chlortetracycline, oxy-tetracycline, tetracycline and semisynthetic tetracyclines, structures, microorganisms, inoculum preparation, production medium, fermentation and recovery methods

Unit IV

Production and application of microbial enzymes. – Amylases, lipases and proteases, uses, microorganisms, inoculum preparation, production medium, fermentation and recovery

Steroid transformations – Substrates, typical structures, microorganisms, inmoc;ulum preparation, 11-hydroxylation, process and recovery.

Principles of vaccine production and types of vaccines

Microbial biopesticides

Microbial products from genetically modified (cloned) organisms eg. Insulin

Paper II MB 452 Microbial Biotechnology (Practicals)

Production of ethanol by flask ferementation, recovery of ethanol by distillation and calculation of fermentation efficiency

Preparation of Beer by Microbial fermentation

Preparation of from grapes/fruits by fermentation

Production of citric acid by fungal fermentation, recovery and estimation

Production of amino acid (Glutamic acid/lysine) by fermentation

Production of amylase by fermentation, recovedry and estimation

Production and estimation of penicillin by flask fermentation

Immobilized bacteria/yeast/enzyme in fermentation

Scale up of fermentation.

Recommended Books

Industrial Microbiology by Casida, LE

Industrial Microbiology by Patel, AH

Industrial Microbiology by Miller, BM and Litsky

Industrial Microbiology by Prescot and Dunn

Microbial Technology by Peppler, JH and Perlman, D.

Biochemistry of Industrial Microorganisms, by Rainbow and Rose

Economic Microbiology by Rose Vol I – V

Microbial Enzymes and Biotechnology by Fogarty WM and Kelly, CT

Comprehensive Biotechnology, All volumes Ed. Murray Moo-Yong

Biotechnoloogy (A text book of industrial Microbiology) Ed. Cruger & Cruger

Advances in Applied Microbiology Ed. Perlman Series of volumes

Laboratory experiments in microbiology by Gopal Reddy et al

M.Sc. (Final) IV Semester Microbiology Paper III MB 403 Microbiology of Food and Environment (4 units x 15 hrs = 60 hrs teaching)

Unit I

Introduction to fermented foods, Microbial products of milk
Microbiology of cheese, butter, yogurt, Microbiology of bread, sauerkraut, idlt
Bacteriological examination of fresh and canned foods
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Food preservation methods, silage. Food poisoning, Mycotoxins
Dairy microbiology - Types of microorganisms in milk, significance of microorganisms in milk, microbni0ological examination of milk, control of microbial flora of milk

Unit II

Microbes and animal interactions – Rumen microbiology, termite microbial communities Microbes in the production of energy from agricultural and domestic wastes Methanogenesis (Biogas) and microbiology of methogenesis

Unit III

Microbial degradation of pesticides and persistence of pesticides Degradation of herbicides, fungicides and insecticides. Microbes and plant interactions – Rhizosphere, Mycorrhizae, Phyllospheree

Unit IV

Microorganisms and water pollution

Water-borne pathogenic microorganisms and their transmission.

Sanitary quality of water. Water pollution due to degradation of organic matter

Aerobic sewage treatment – Oxidation ponds, trickling filters, activated sludge treatment

Anaerobic sewage treatment – Septic tank

Microorganisms in air and their importance (brief account)

Paper III MB 453 Microbiology of Food and Environment (Practicals)

Microbiological examination of fresh and canned foods and mushrooms

Microbiological examination of milk and milk products

Microbiological quality testing dof milk (MBRT test)

Isolation and cultivation of anaerobic microbes from rumen, and termites

Isolation and observation for phillosphere microflore

Isolation and observation for rhizosphere microflore

Observation for mycorrhizae

Effect of pesticides on microbial activity

Estimation of BOD

Testing for microbial sanitary quality of water (coliform test)

Isolation and analysis of mycotoxins
Isolation and observation of air microsflora

Recommended books

Food Microbiology by Frazier
Microbial Ecology – A conceptual approach by Lynch and Poole
Basic food microsbiologoy (Abridged edition) by George J. Banwart
Waste water microbiology by Bitton, G.
Waster water treatment – Biological and chemical process by Henze, M.
Soil Microbiology by Alexander Martin
Soil Microorganisms and Plant growth by NS Subba RAo
Laboratory experiments in microbiology by Gopal Reddy et al

M.Sc. (Final) IV Semester Microbiology Paper IV MB 404 Medical Virology and Parasitology (4 units x 15 hrs = 60 hrs teaching)

Unit I

Diagnostic virology – Cultivation of pathogenic viruses in lab animals and tissue culture Identification of pathogenic viruses and establishment of viral etiology Air borne viral infections (detailed study) – Influenza virus, Rhino virus, Rubella virus, Adeno virus (type 2), Mumps virus and Measclesvirus

Unit II

Detailed study of viruses transmitted by water - Hepatitis (HAV), Rmk, Polio myelitis Detailed study of viruses transmitted by Zoonosis - Rabies, Japanese encephalitis

Unit III

Detailed study of contact and sexually transmitted viral diseases – Small pox, Herpes (Herpes simplex virus), Hepatitis viruses and their diseases Acquired immunodeficiency syndrome (AIDS)

Unit IV

Malaria, Amoebiasis, Helmithic infections (Round worms, Hookd worms), Medical Mycology – Dermatomycosis, Systemic mycosis

IV MB 454 Medical Virology and Parasitology (Practicals)

Cultivation of viruses - Egg inoculation (CAM, Allantoic, Amniotic route inoculation) Tissue culture techniques

Animal inoculation techniques
Microscopic studies of viruses infected materials
Examination of pathogenic fungi
Examination of stool for helminthes and Amoeba
Examination of blood smear by Leishman stain for medical parasites
Rota viral RNA (ds RNA) analysis
Immunodiagnosis - ELISA tests

Recommended Books

Review of medical microbiology by Jawitz et al Medical laboratory Manual for tropical countries Vol I & II by Monica Cheesbrough Text Book of Microbiology by Ananthanarayanan and Jayaram Paniker Viral and Ricketsial infections of Man by Horsfall and Jam Text book of Virology by Rhodes and Van Royan Virological Procedures by Mitchal hasking Virologoy by Wilson and Topley